Cod: 5.4006

ENVIRONMENTAL QUALITY AND FOOD SAFETY: BIOINDICATORS OF PERSISTENT ORGANIC POLLUTANTS EXPOSURE

<u>G. Diletti¹</u>, G. Brambilla², L. Candela³, R. Ceci¹, A. Conte¹, S.P. De Filippis², A. Di Sandro³, S. Menotta⁴, A. Ubaldi⁵, G. Scortichini⁶

¹*Istituto Zooprofilattico Sperimentale dell'Abruzzo e del Molise, Teramo, Italy*

²*Istituto Superiore di Sanità, Rome, Italy*

³*Ministero della Salute, Rome, Italy*

⁴Istituto Zooprofilattico Sperimentale della Lombardia e dell'Emilia-Romagna, Bologna, Italy

⁵*Istituto Zooprofilattico Sperimentale del Lazio e della Toscana, Rome, Italy*

⁶Istituto Zooprofilattico Sperimentale dell'Umbria e delle Marche, Perugia, Italy

Introduction

Food of animal origin represents the main source of exposure for the general population to persistent organic pollutants (POPs). The relationship between environmental quality and POPs bioaccumulation and bioconcentration is consolidated, as well as the use of animal species as bioindicators of contaminant exposure¹. In 2009-2010, a pilot study was conducted in Italy to evaluate sheep/goat and honeybees as potential bioindicators of environmental contamination. Dioxins and furans (PCDDs/PCDFs), dioxin-like and non dioxin-like polychlorinated biphenyls (DL-PCBs/NDL-PCBs), polybrominated diphenyl ethers (PBDEs), perfluoroalkyl acids (PFAAs), pentachlorobenzene (PeCB), hexachlorobenzene (HCB) and pentachlorophenol (PCP) were selected as POPs to be monitored in sheep/goat milk and honey. The samples were collected from areas affected by the presence of POP emission sources, and from control areas, to highlight differences in the levels of contamination.

Materials and methods

Sampling plan

In order to provide representative bioindicators, milk from sedentary sheep or goat flocks and wildflower honey were sampled. Indeed, sedentary animals better reflect the environmental quality, while wildflower honeys are produced in a longer period in comparison to monofloral ones. The samples were taken: a) nearby pollutants emission sources or contaminated areas already identified, marked as exposed (E) zones; b) control sites in remote areas, such as national or regional natural parks, marked as non-exposed (NE) zones. Samples were collected from 16 Italian regions: a) for milk, 15 samples were from E-zones, 13 from NE-zones; b) for honey, 15 sample were from E-zones, 12 from NE-zones (Table 1).

Analytical methods

For PBDEs, 9 congeners (BDE-28, BDE-47, BDE-66, BDE-85, BDE-99, BDE-100, BDE-153, BDE-154, BDE-183) were analyzed using ¹³C labelled internal standards. Milk samples were extracted according to AOAC 905.02, then subjected to acidic treatment and purification on a multi-layer silica column (silica gel and acidic silica). Honey samples were dissolved in water and extracted with n-hexane, while the clean-up procedure was the same as for milk. Instrumental analysis was performed by isotope dilution gas-chromatography – high resolution mass spectrometry (GC-HRMS) at resolution > 10,000 using a DB-5 MS J&W (60 m x 0.25 mm x 0.10 μ m) column. The sum of nine PBDE congeners was calculated as upper-bound (UB) concentration.

For PCDDs/PCDFs and DL-PCBs/NDL-PCBs, methods based on EPA 1613 rev. B and EPA 1668 rev. B were used, respectively. Milk samples were lyophilized and extracted with toluene by accelerated solvent extraction (ASE). Honey samples were dissolved in water and extracted with dichloromethane. In both cases, the extract was first purified on Extrelut[®] column acidified with sulphuric acid, and then by Power-Prep[®] using multi-layer silica gel, alumina and carbon columns. The PCDDs/PCDFs were separated from DL-PCBs/NDL-PCBs. Instrumental analysis was performed by GC-HRMS at resolution > 10,000 using a DB-5 MS J&W (60 m x 0.25 mm x 0.25 µm) column. PCDDs/PCDFs and DL-PCBs were expressed as UB concentration using WHO₂₀₀₅-TEF scale. NDL-PCBs analytical sum was also calculated as UB value. For PCP, two different analytical procedures were applied to milk and honey samples, both using ¹³C labelled PCP as internal standard. Milk samples were extracted following the same procedure used for PCDDs/PCDFs analysis, then purified by solid-phase extraction (SPE) on a silica-gel cartridge. The final solution was analyzed by liquid-chromatography – tandem mass spectrometry (LC-MS/MS) using an XTerra C18 column (150 x 2.1 mm, 3 µm) as stationary phase, and a water/acetonitrile solution containing formic acid as mobile phase. Honey samples were extracted with water/acetonitrile, then adding magnesium sulphate, sodium chloride, and buffering salts according to the QuEChERS procedure. PCP was determined by gas chromatography – ion trap mass spectrometric detection (GC – ITD) after acetylation, using a RTX-Sil MS (30 m x 0.25 mm x 0.25 µm) column.

PFOS/PFOA were extracted with acetonitrile, previously adding their respective ¹³C labelled internal standards and dissolving honey in water. The extract was purified onto an Oasis WAX 150 mg cartridge. The instrumental analysis was performed by LC-MS/MS using a Symmetry C18 (150 mm x 2.1 mm, 3 μ m) column and water/methanol (both containing 0.01% formic acid) as mobile phase.

PeCB/HCB in milk samples were analyzed by the same procedure used for PBDEs, previously adding a ¹³C labelled HCB internal standard. Honey samples were dissolved in water and extracted with n-hexane, then purified on an Extrelut[®] column impregnated with sulphuric acid. The analytical determination was accomplished by GC-HRMS using a HT8-PCB (60 m x 0.25 mm x 0.25 µm) column.

Results and discussion

Milk samples

The results are summarised in Table 2. The Mann-Whitney U non-parametric test for independent groups was used to compare the results for E/NE-zones. The statistical evaluation was not performed for PCP and PFOA due to the high frequency of results below the limit of quantification.

Statistically significant differences (p = 0.05) between zones were found:

• for PCDDs/PCDFs (max 1.88, Lazio E-zone - min 0.04, Tuscany NE-zone, pg WHO₂₀₀₅-TEQ/g fat);

• for DL-PCBs (max 3.33, Lombardy E-zone – min 0.06 Lazio NE-zone, pg WHO₂₀₀₅-TEQ/g fat).

Four samples from E-zones were above the PCDDs/PCDFs and DL-PCBs action levels designed to prompt competent authorities and operators to identify a source of contamination and to take measures for its reduction or elimination. In particular, the action level was exceeded for PCDDs/PCDFs in Colleferro (Lazio), and for DL-PCBs in Val Trompia (Lombardy), Milazzo (Sicily) and Crotone (Calabria). These sites are characterized by the presence of industrial plants, some of them have been also classified as contaminated sites of national interest (Table 1).

PBDE sum was in the range 146-920 pg/g fat with predominant congeners in the order 47 > 99 > 100 > 154 reflecting past use of technical mixtures constituted of penta- and hexa-bromodiphenylethers as flame retardants². Regarding PFAAs, the maximum values for PFOS (0.26 ng/g, Sardinia E-zone) and PFOA (0.02 ng/g, Umbria NE-zone) were below the mean values reported in the literature³.

For HCB, the obtained values were in the range 1.69 - 9.27 ng/g fat (EU maximum level 0.01 mg/kg fat), while for PeCB concentrations between 0.05 and 0.57 ng/g fat were recorded were recorded. All samples were negative for PCP (< 5 ng/g).

Honey samples

The Mann-Whitney U test could not be performed for honey samples because most of results were below the LOQ values. This matrix seemed not suitable for POP levels monitoring, nevertheless useful data were obtained for heavy metals and trace elements as a part of the study not discussed here. In conclusion, honey contamination appears mainly attributable to beekeeping practices (e.g. acaricides and antibiotics use) rather than environmental contaminants, excluding heavy metals and trace elements.

Acknowledgements

The authors wish to acknowledge the Italian Ministry of Health for the financial support of this study.

References

1. Scortichini, G., Amorena, M., Brambilla, G., Ceci R., Chessa, G., Diletti, G., Esposito, M., Esposito, V., and Nardelli, V. (2016) Small Rum Res 135, 66-74.

2. Schecter, A., Haffner, D., Colacino, J., Patel, K., Päpke, O., Opel, M., and Birnbaum, L. (2010) Environ Health Perspect 118, 357-262.

3. Hlouskova, V., Hradkova, P., Poustka, J., Brambilla, G., De Filipps, S. P., D'Hollander, W., Bervoets, L., Herzke, D., Huber, S., de Voogt, P., Pulkrabova, J. (2013) Food Addit Contam Part A Chem Anal Control Expo Risk Assess 30 (11), 1918-1932.

Region	Contaminated area (milk – honey)	Control area (milk – honey)		
Sardinia	Fiume Santo (SS) ¹ – Sulcis Iglesiente Guspinese (CI) ²	Sassari – Lula (NU)		
Sicily	Milazzo (ME) ²	Nebrodi Regional Park (ME)		
Tuscany*	Le Strillaie $(GR)^2$	Grosseto		
Marche	Basso bacino del fiume Chienti $(MC)^2$	Gola della Rossa Regional Park AN)		
Lazio	Colleferro (RM) ³ – Segni (RM) ⁶	Monti Simbruini Regional Park (RM)		
Abruzzo	Bussi Popoli(PE) ²	Gran Sasso Monti della Laga National Park (TE)		
Campania	Valle del Sarno $(SA)^2$	Cilento National Park (SA)		
Trentino	Valsugana (TN) ⁴			
Molise	Campobasso Guglionesi II (CB) ²	Abruzzo National Park (IS)		
Umbria	Terni Papigno (TR) ²	Monti Sibillini National Park (PG)		
Basilicata	Val Basento $(MT)^2$	Pollino National Park (MT)		
Lombardy	Val Trompia (BS) ⁴			
Apulia	Taranto ²	Gargano National Park (FG)		
Emilia- Romagna	Formigine (MO) ⁵	Appennino Tosco Emiliano National Park (BO)		
Calabria	Crotone ²	Sila National Park (CS)		
Piedmont**	Carisio $(VC)^7$			

Table 1. Sampling sites for sheep/goat milk and honey

*Only milk samples taken; **Only honey samples taken

¹Thermal power plant; ²Contaminated site of national interest; ³Metal industries, organochlorine pesticides production, cement kilns, waste incinerators; ⁴Steel plants; ⁵Ceramic factories; ⁶Cement kiln; ⁷Aluminum smelter.

Table 2. Analytical results for sheep/goat milk samples											
Region	Group	PBDEs ^a pg/g fat	PCDDs/PCDFs pg WHO ₂₀₀₅ - TEQ/g fat	DL-PCBs pg WHO ₂₀₀₅ - TEQ/g fat	NDL-PCBs ng/g fat	PeCB ng/g fat	HCB ng/g fat	PFOS ng/g			
Sardinia	E-zone	257	0.17	0.49	2.4	0.379	3.929	0.2557			
	NE-zone	419	0.09	0.59	2.4	0.134	2.620	0.0040			
Sicily	E-zone	511	0.72	2.91	6.4	0.166	2.801	0.0355			
	NE-zone	363	0.29	0.28	2.4	< 0.086	2.492	0.0037			
Tuscany	E-zone	442	0.61	0.63	2.4	0.354	3.903	0.0033			
	NE-zone	292	0.04	0.28	2.4	< 0.082	3.255	0.0026			
Marche	E-zone	276	0.35	0.76	4.1	0.267	4.983	0.1365			
	NE-zone	321	0.69	0.67	3.5	0.084	3.836	0.0018			
Lazio	E-zone	379	1.88	1.30	2.7	0.177	5.104	0.0041			
	NE-zone	376	0.20	0.06	2.4	0.286	4.150	0.0045			
Abruzzo	E-zone	146	0.11	0.66	2.6	0.193	3.388	0.0054			
	NE-zone	335	0.10	0.68	4.4	0.195	4.896	0.0148			
Campania	E-zone	432	0.71	1.31	2.4	0.143	3.548	0.0078			
	NE-zone	351	0.31	0.95	5.3	0.242	2.291	0.0049			
Trentino	E-zone	518	0.51	1.71	4.6	0.538	9.270	0.0160			
Molise	E-zone	567	0.11	0.80	2.8	< 0.235	2.501	0.0053			
	NE-zone	298	0.11	0.72	2.7	< 0.113	1.686	0.0054			
Umbria	E-zone	244	0.73	1.63	5.0	0.145	3.057	0.0034			
	NE-zone	320	0.11	0.67	3.0	0.444	4.600	0.0039			
Basilicata	E-zone	423	1.17	0.59	3.7	0.066	2.952	0.0033			
	NE-zone	274	0.19	0.34	3.5	< 0.069	3.224	< 0.0056			
Lombardy	E-zone	920	0.30	3.33	13.3	0.239	3.098	0.0121			
Amilia	E-zone	277	0.53	1.74	9.1	0.570	5.008	0.0137			

T

^a Sum of BDE-28, BDE-47, BDE-66, BDE-85, BDE-99, BDE-100, BDE-153, BDE-154, BDE-183

0.28

0.75

0.09

0.10

0.12

^b Not determined due to the signal from procedural blanks

NE-zone

E-zone

NE-zone

E-zone

NE-zone

398

645

488

457

327

Apulia

Calabria

Emilia-Romagna

0.54

0.94

0.67

2.64

1.91

2.8

3.5

3.1

10.3

5.4

0.080

< 0.201

0.059

0.046

< 0.170

1.809

1.926

2.374

3.765

3.687

0.0124

< 0.0057

0.0388

0.0056

0.0028

PFOA

ng/g

< 0.0090

< 0.0092

ND

 ND^{b} < 0.0080

< 0.0077

< 0.0086

< 0.0082

< 0.0082

< 0.0079

< 0.0085

< 0.0087

< 0.0085

< 0.0084

< 0.0087

< 0.0085

< 0.0084

< 0.0084

0.0188

< 0.0090

ND^b

< 0.0081

< 0.0083

< 0.0083

ND

< 0.0085

< 0.0101

< 0.0096