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# ALTERNATIVE SULPHURIC ACID PRE-TREATMENT AND SILICA BASED SOLVENT CHANGE DEMONSTRATED FOR LARD AND FISH OIL ANALYSED BY EROD-BIOASSAY

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## Introduction

Bioassays for routine application should be simple and robust to use. Some samples tend to plug<sup>1</sup> the clean up column. Aim of the study was to develop a solution without further equipment or chemicals. Concepts of column clean up with and without pre-treatment were compared.

## Materials and methods

Aliquots of lard and bovine fat were spiked with 20  $\mu$ l 2,3,7,8-TCDD/PCB-126 (1/10 w/w) in DMSO. Every subsample with its spike was freshly prepared 6fold. Fish oil was tested without spike. Original samples were as well analysed via HRGC/HRMS. 2,3,7,8-TCDD and PCB-126 were from LGC-Standards. The quality of all other chemicals, the preparation of solutions and the EROD-bioassay (PBS/dispense) using rat hepatoma cells (H4IIEC/T3) criteria were described before<sup>2,3,4,5</sup>. The clean-up procedure depended on the tested amount of fat, see table 1.

Basic column: The clean up for 1 g bovine fat<sup>3,6</sup> was scaled up for 2 g lard.

## "Dry" sulphuric acid pre-treatment:

3 g lard were spiked and solved in 5 ml n-hexane. 25 ml acidic silica (44 w%  $H_2SO_4$ ) were added and mixed with the fat. This step increased the recovery of spikes due to less displacement of polar DMSO by nonpolar n-hexane (data not shown). After two to five minutes 15 ml n-hexane were added and agitated. Some of the solvent evaporated so the consistence of the mixture allowed quickly to pour the whole blend on top of the corresponding clean up column.

## Silica based solvent change

For fish oil a coupling of fat extraction and acidic treatment preceding the cleanup column was tested. Lipids were extracted by adding Na<sub>2</sub>SO<sub>4</sub> and eluting with 200 ml n-hexane/acetone  $2/1 (v/v)^{3,4,7}$ . Fat extracts were reduced, transferred with 5 ml n-hexane/acetone 2/1 (v/v) to 10 ml heated silica and mixed. Solvents were removed by evaporation. The dry blend was mixed with 15 ml acidic silica (44 w% H<sub>2</sub>SO<sub>4</sub>). The mixture was moistened by 15 to 20 ml n-hexane, mixed again and poured onto a clean up column as described for 3 g lard.

## **Results and discussion**

The comparison of data for bovine fat to the larger column for 2 g lard is shown in table 2. The curve parameters show similar values. Most authors<sup>2,6,8</sup> use silica clean up of comparable acid to fat ratio. Our results for basic columns fit well into the scheme. They lead to the assumption of similar dose-response-curves for fat from different mammals. Further data must be gained to verify which kinds of fat can be gathered to matrix groups in routine analysis.

Figure 1 compares 2 g lard cleaned up on basic columns and 3 g lard with preceding acidic silica treatment respectively. Samples of identical concentration (0.1; 1.1; 2.1 pg BEQ/g fat) show no significant difference (p=0.05) between the concepts. The curve parameters are almost identical. Other authors use acidic pre-treatment with considerable amounts of n-hexane<sup>1,9</sup>. Liquid sulphuric acid<sup>1</sup> or loose bulk acidic silica<sup>9</sup> are used. The advantages of the concepts are discussed in table 3.

In a following step the concept of "dry" silica pre-treatment was tested for fish oil (1 g each) and for some samples coupled to fat extraction as described above. No significant differences (p=0.05) could be observed between extracts obtained by different extraction and clean up combinations, s. table 4.

By using "dry" silica based sulphuric acidic pre-treatment samples with lipophilic compounds not soluble in n-hexane can be addressed. Further it is possible to pre-treat samples that tend to plug the clean up column like rancid oil. A transfer of polar lipids to the cleanup is possible without acetone while washing<sup>1</sup> is avoided. No change in solvent composition or devices used so far was necessary. Depending on the amount of fat the concept can be scale up. When working with DMSO-spikes the use of glassware is strongly recommended. For naturally contaminated fish oil vessels from polypropylene obtained no negative effect.

#### **References:**

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Alternative sulphuric acid pre-treatment and silica based solvent change demonstrated for lard and fish oil analysed by EROD-bioassay

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## Abstract attach: Tables and figures

**Table 1**: Set up of columns for clean-up. Silica was added by measuring spoon, with8 ml representing 5 to 5.5 g

Column set up	$\leq$ 1 g fat <sup>3,6</sup> $\leq$ 2 g fat		≤ 3 g fat	
top	3 g Na <sub>2</sub> SO <sub>4</sub> 3 g Na <sub>2</sub> SO <sub>4</sub>		Mixture from pre-treatment	
H <sub>2</sub> SO, on silica	8 ml 22 w% H <sub>2</sub> SO <sub>4</sub>	16 ml 22 w% H <sub>2</sub> SO <sub>4</sub>	16 ml 44 w%	
$\Pi_2 S O_4 \text{ OIT SIIICA}$	8 ml 33 w% H <sub>2</sub> SO <sub>4</sub>	16 ml 44 w% H <sub>2</sub> SO <sub>4</sub>	$H_2SO_4$	
bottom	3 g Na <sub>2</sub> SO <sub>4</sub> on glass wool			
Conditioning of column before addition of samples				
n-hexane	40 ml 100 ml		40 ml	
Application of sample	solved in 2 to	pouring of mixture		
Elution of sample after rinsing of the sample vessels				
n-hexane	2 x 5 ml + 28 ml 2 x 5 ml + 140 ml 2 x 5 ml + 65 ml			

**Table 2**: Validation of spiked fat, each concentration was extracted 6fold and measured via EROD-bioassay. Mean values of bioassay with coefficients of variation (CV), curve parameters and HRGC/HRMS results are shown.

Bovine fat 1 g, basic column <sup>3</sup>		Lard, 2 g fat, basic column		
HRGC/HRMS	EROD	HRGC/HRMS	EROD	
pg WHO-TEQ <sup>5</sup> /g fat	pg BEQ/g fat	pg WHO-TEQ⁵/g fat	pg BEQ/g fat	
( $\Sigma$ PCDD/F +dl PCB)	± CV %	( $\Sigma$ PCDD/F +dl PCB)	± CV %	
-	-	0.14	0.10 ± 56.6 %	
-	-	0.89	0.73 ± 17.6 %	
-	-	1.14	1.20 ± 11.0 %	
2.88	3.01 ±8.60 %	1.64	1.58 ± 11.0 %	
4.84	5.15 ±18.0 %	2.14	2.24 ± 16.0 %	
6.84	6.85 ±8.40 %	3.14	3.58 ± 14.3 %	
8.88	9.78 ±15.1 %	4.14	4.16 ± 13.5 %	
Parameter of	y = 1.15x - 0.49	Parameter of	y = 1.07x - 0.09	
linear curve	$R^2 = 0.95$	linear curve	$R^2 = 0.94$	



**Figure 1:** Validation of 2 g lard on basic columns and for 3 g lard with acidic silica pre-treatment. Each concentration was extracted 6fold and measured via EROD-bioassay. One point represents the mean of three wells. Results are based on WHO-TEF<sup>5</sup>. Confidence and prediction intervals for 2 g lard, curve parameters for linear fit and coefficients of determination ( $R^2$ ) are shown.

Concept	<b>A</b> makura <sup>1</sup>	US EPA <sup>9</sup>	this study	
samplo	30-50 g	5 g	≤ 3 g	
Sample	fast food	soil/sediment	fat	
visible n-hexane phase	$\checkmark$	$\checkmark$	-	
ovidation	30-50 ml	H₂SO₄ on silica	25 ml 44 w%	
UXIDATION	$H_2SO_4$ conc.	small amount	H <sub>2</sub> SO <sub>4</sub> on silica	
standard mixing glass ware	1	tight and inert	1	
standard mixing glass ware	•	sealing	•	
scaling possible	$\checkmark$	$\checkmark$	$\checkmark$	
optical control	clear n-hexane-phase		browning	
load slurry* on column	-	$\checkmark$	$\checkmark$	
specific needs	washing steps prevention of charring	shaker	-	

**Table 3**: Comparison of acidic pre-treatment concepts

\* sample mix with acidic silica after oxidation

Table 1. Compandon of amoronic oxtraction and oldari up conteniou for righter	Table 4: Com	parison of differer	nt extraction and clean	-up schemes for 1	g fish oil
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Group	Number of	Fat	Sulphuric acid	pg BEQ/g fat
Group	samples n	Extraction	Pre-treatment	± CV %
A - control	16	-	-	1.62 ± 23.9%
В	12	-	$\checkmark$	1.64 ± 9.5%
С	4	$\checkmark$	$\checkmark$	1.66 ± 11.4%