

# POLYCHLORINATED DIBENZO-*p*-DIOXINS AND DIBENZO-FURANS IN LIVER OF PIGLET ACCORDING TO DISEASES

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## Introduction

Persistent Organic Pollutants (POPs) are chemical substances that persist in the environment, bioaccumulate through the food web and pose a risk of causing adverse effects to human health and the environment<sup>1</sup>. Polychlorinated dibenzo-*p*-dioxins (PCDDs) and dibenzofurans (PCDFs) are found in a variety of foods. Ninety per cent of human exposure to POPs comes from food<sup>2</sup>. The major food sources are fish and seafood and dairy products<sup>1</sup>. A number of studies also reported high levels of PCDD/Fs in animals and public concern about PCDD and PCDF levels in animals has been raised<sup>3-4</sup>.

Globally, there has been increase in the breeding of pig more than cattle. Koreans are consuming pork more than beef and chicken<sup>5</sup>. Pigs contaminated by POPs are a serious public health issue<sup>6</sup>.

Porcine reproductive and respiratory syndrome (PRRS) has become endemic and now is an important cause of pneumonia in 3.24-week-old pigs in swine producing countries around the world. Several research groups have demonstrated increased severity of clinical disease in animals coinfecting with PRRS virus (PRRSV) and different bacteria, including *Streptococcus suis*, *Haemophilus parasuis*, *Salmonella choleraesuis* and *Mycoplasma hyopneumoniae*<sup>7</sup>. Exposure of PRRSV-infected pigs to different *S. suis* strains resulted in different clinical manifestations and severity of disease, indicating that the strain of bacteria can influence the clinical disease induced in a coinfection model<sup>8</sup>. However, the mechanisms of the interaction between PRRSV and bacterial infections remains unclear<sup>7</sup>.

The purpose of this study was to evaluate interrelation between the levels of PCDD/Fs in liver of piglets and diseases of piglet.

## Materials and methods

### 1) Standards and Reagents

The following <sup>13</sup>C-labeled and <sup>37</sup>Cl-labeled PCDD/Fs solutions for EPA 1613 were purchased from Wellington laboratories Inc.(Ontario, Canada). For the quantification of low concentrations of PCDD/Fs, calibration standards CS1/2 through CS4 were used. All organic solvents were of ultra-residue grade analysis (J.T Baker, Philipsburg, NJ, USA). Anhydrous sodium sulfate and sulfuric acid from E. Merck (Darmstadt, Germany) were used.

### 2) Samples

It was consisted two experiments that analysis of PCDD/Fs in pig liver according to diseases samples and analysis of PCDD/Fs in liver. In first, Twenty 3-4 week-old piglets were assigned to 4 groups of 3 piglets each. The diseases of piglet were Bacillus nature diseases(*Salmonella* spp and *Streptococcus suis*) and Virus diseases (PRRS:Porcine Reproductive and Respiratory Syndrome, Lung), PRRS(Lymph, Intestine).

In second, Ten 7 day-old piglets were assigned to 2 groups of 5 piglets each. The diseases of piglet were Virus diseases(North America type, European type).

### 3) Analytical condition

The preparation of samples(Fig. 1) was based on the EPA method 1613 with automated sample cleanup for trace analysis(PowerPrep<sup>TM</sup>, FMS).

The instrumental analysis was performed on a JEOL(Akishima, Tokyo, Japan) MStation JMS-700D high resolution mass spectrometer (B/E configuration) equipped with a Agilent (Palo Alto, CA, USA) 6890 Plus gas chromatograph. The capillary GC column 30 m•0.32 mm, coated with a DB-5MS stationary phase (film thickness 0.25um) was used. Samples were injected in splitless mode at an injector temperature of 280 °C and at an initial column temperature of 160°C. After 1 min., the temperature was ramped at 20°C/min to 200°C, at

5°C/min up to 235°C, and at 3°C/min up to 310°C. The latter temperature was held for 3 min. The ion source was operated at 260 C, the electron energy was 38 eV. The mass spectrometer was tuned to a mass resolution of 10,000. When signal to noise ratio (S=N) for a given peak was lower than 3, congeners were recorded as nondetected (n.d.) for quantification. Quantification was carried out Tetra- through Octa-Chlorinated Dioxins and Furans by the isotopic dilution method.

## Results and Discussion

### 1) The comparison of bacteria diseases and virus disease

All 17 of the 2,3,7,8-chloro-substituted dibenzo-p-dioxins and dibenzofurans included in the WHO methodology were analyzed. No detectable concentrations of PCDD/Fs were observed in piglet liver according to the diseases of Salmonella spp. and Streptococcus. In piglet liver according to the diseases of PRRS(lung), all 17 of the 2,3,7,8-chloro-substituted OCDD and dibenzofurans were not detected on two samples. Only 2,3,7,8-chloro-substituted OCDD were detected on one sample. In the diseases of PRRS(lymph and intestine), 1,2,3,7,8-pentachlorodibenzo-p-dioxin (PCDD), the most potent dioxins, was observed in one sample. 2,3,7,8-TCDF, 1,2,3,4,7,8-HxCDD, 1,2,3,4,6,7,8-HpCDF, 1,2,3,4,7,8,9-HpCDF were detected(Table 1). 1,2,3,4,6,7,8-HpCDF was detected in 3 samples. Salmonella spp. and Streptococcus is Bacillus nature diseases. PRRS is Virus diseases. The concentrations of PCDD/Fs in piglet liver from Virus diseases was higher than the concentrations from Bacillus nature diseases.

The recoveries of sampling and clean-up standards for dioxin congener which indicated the loss of the procedure. The recoveries of sampling and clean-up standards ranged 73-110% which were under the required range. Method blanks have been also run.

### 2) The comparison of PRRS in north American type and European type

The PCDD/Fs in piglet liver from PRRS of North America type were detected more than from PRRS of Europe(Table 2). The concentrations of PCDD/Fs in piglet liver from PRRS of North American type were observed higher than from PRRS of European type.

As a result of congener specific analysis, the concentrations determined in this study are in agreement with results published by reports<sup>9</sup>. The concentrations of PCDD/Fs from the piglet of Virus disease were more than from Bacillus nature disease.

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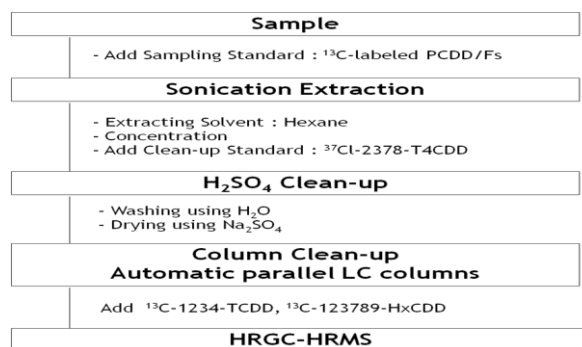


Fig. 1 Schematic diagram for analytical procedure

Table 1. The concentrations of PCDD/Fs in piglet liver according to diseases samples

Compounds	(pgWHO-TEQ/g)											
	Salmonella spp. and Streptococcus suis			Streptococcus suis			PRRS (Lymph, Intestine)			PRRS(Lung)		
	1	2	3	4	5	6	7	8	9	10	11	12
2,3,7,8-TCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8-PeCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	0.4300	N.D	N.D	N.D	N.D
1,2,3,4,7,8-HxCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	0.0055	N.D	N.D	N.D	N.D
1,2,3,6,7,8-HxCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8,9-HxCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,4,6,7,8-HpCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
OCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
2,3,7,8-TCDF	N.D	N.D	N.D	N.D	N.D	N.D	0.0550	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8-PeCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
2,3,4,7,8-PeCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,4,7,8-HxCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,6,7,8-HxCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8,9-HxCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
2,3,4,6,7,8-HxCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,4,6,7,8-HpCDF	N.D	N.D	N.D	N.D	N.D	N.D	0.0067	0.0045	0.0042	N.D	N.D	N.D
1,2,3,4,7,8,9-HpCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	0.0049	N.D	N.D	N.D	N.D
OCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	0.0002
ΣPCDD/F	N.D	N.D	N.D	N.D	N.D	N.D	0.0617	0.4449	0.0042	N.D	N.D	0.0002

N.D: not detected

**Table 1. The concentration of PCDD/Fs in pig liver according to Bacillus nature diseases**

(pgWHO-TEQ/g)

Compounds	PRRs of North American type					PRRs of European type				
	NA-1	NA-3	NA-5	NA-7	NA-15	EU-1	EU-3	EU-5	EU-7	EU-10
2,3,7,8-TCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8-PeCDD	N.D	N.D	N.D	N.D	0.2095	N.D	N.D	N.D	N.D	N.D
1,2,3,4,7,8-HxCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,6,7,8-HxCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8,9-HxCDD	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,4,6,7,8-HpCDD	N.D	0.0011	N.D	N.D	0.0028	N.D	N.D	N.D	N.D	N.D
OCDD	N.D	N.D	0.0001	N.D	0.0001	0.0003	0.0002	0.0001	N.D	N.D
2,3,7,8-TCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8-PeCDF	0.0065	0.0031	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
2,3,4,7,8-PeCDF	0.0650	0.0437	N.D	N.D	0.1396	N.D	N.D	N.D	N.D	N.D
1,2,3,4,7,8-HxCDF	0.0208	N.D	N.D	N.D	0.0349	N.D	N.D	N.D	N.D	N.D
1,2,3,6,7,8-HxCDF	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D	N.D
1,2,3,7,8,9-HxCDF	N.D	N.D	N.D	0.0183	N.D	N.D	N.D	N.D	N.D	N.D
2,3,4,6,7,8-HxCDF	N.D	0.005	N.D	N.D	0.0349	N.D	N.D	N.D	N.D	N.D
1,2,3,4,6,7,8-HpCDF	0.0036	N.D	0.0028	N.D	N.D	N.D	N.D	0.004	N.D	N.D
1,2,3,4,7,8,9-HpCDF	N.D	0.001	N.D	N.D	0.0063	N.D	N.D	N.D	N.D	N.D
OCDF	N.D	N.D	N.D	N.D	0.0001	N.D	N.D	N.D	N.D	N.D
ΣPCDD/F	0.0959	0.0539	0.0029	0.0183	0.4282	0.0003	0.0002	0.0041	0	0

N.D: not detected