

DETERMINATION OF FLUOROTELOMER ALCOHOLS AND PERFLUOROALKYL CARBOXYLATES IN AIR ENVIRONMENT IN THAILAND

Kongpran J^{1*}, Tanaka S¹, Kunacheva C², Fujii S¹, Sakui N³, Boontanon SK⁴, Suzuki Y¹, Harada H¹

¹GSGES, Kyoto University, Yoshida Campus, Sakyo-ku, Kyoto, Japan; ²Nanyang Environment & Water Research Institute, Nanyang Technological University, 1 Cleantech Loop, CleanTech One, Singapore; ³Agilent Technologies Japan, Ltd., Hachioji, Tokyo, Japan; ⁴Faculty of Engineering, Mahidol University, Nakhon Pathom, Thailand

Introduction

Perfluorinated compounds (PFCs) are used as processing additives, during fluoropolymer productions, and surfactants in consumer products for over 50 years¹. Perfluoroalkyl carboxylates (PFCAs) and sulfonates (PFASs), which are the most concerned groups of PFCs, are known to be persistent, bioaccumulative and toxic to human and animals². PFCAs and PFASs are highly water soluble but less volatile³, and they have been detected in water environment worldwide⁴⁻⁵. PFCAs are not found in significant quantities as a vapor but they can be associated with particulate matter in the air⁶. Precursor compounds such as fluorotelomer alcohols (FTOHs), which can be degraded to PFCAs⁷⁻⁸ and are more volatile, were detected in the air not only in developed countries⁹⁻¹³ but also in a global scale along Atlantic and Southern oceans¹⁴. Information on contamination of FTOHs and PFCAs in air environment has been developed but mainly in developed countries, whereas very few studies have been conducted in developing countries including Thailand. PFCs contamination was observed in water and wastewater in Thailand¹⁵⁻¹⁶ but the contamination in the air has never been studied before. This study is the first investigation of FTOHs and PFCAs in outdoor and indoor air in Thailand, and the concentrations were compared with those in previous studies. In addition, composition profiles and particle-gas partitioning of the target compounds were examined in order to interpret their possible sources and behavior in the atmosphere.

Materials and methods

Sample Collection: Survey was conducted in the Central and Eastern parts of Thailand. Various outdoor and indoor areas, which are suspected to be contaminated by FTOHs and PFCAs, were selected as sampling sites. Detailed characteristics of the sites; wastewater treatment plants (WWTPs) and residential areas for outdoor air, and the places for indoor air, are demonstrated in **Table 1**. Samples were collected during 10 September - 29 October 2012 by using a high volume air sampler (Sibata, Japan) at the flow rate of approximately 500 L/min for 24 h (~720 m³). This study adapted three types of sampling media that comprised of glass fiber filters (GFFs, Advantec®, Japan), polyurethane foams (PUFs, Sibata, Japan) and activated carbon fiber felts (ACFs, Sibata, Japan). All the sampling media were pre-cleaned by soaking into ethyl acetate for 2 times and methanol for 3 times. One set of the sampling media (GFF and PUF-ACF-PUF) was used to collect the target compounds at each sampling site. Particle phase fractions of the target compounds were collected on GFF, while gas phase fractions were trapped in a column of PUF-ACF-PUF. Another set of the media was also prepared at four sampling sites (1, 2, 7 and 9) as a field blank to examine contamination during all of the procedures. The field blank set was just placed close to the sampling points without installation in the vacuum pump for sampling.

Sample Preparation: For particle phase FTOHs extraction, a half of GFF was soaked in 10 mL methanol and shaken for 1 hour. The extraction process was repeated for 4 cycles (~40 mL in total). For gas phase FTOHs extraction, PUFs and ACF were separately soaked and shaken in methanol. The extraction was done for 4 cycles and extracts were combined (~900 mL in total). Mass-labeled FTOH (¹³C₂ 8:2 FTOH) was spiked into the sampling media before extraction to determine the recovery. Extracts of particle and gas phases FTOHs were separately evaporated to 3-5 mL by using a rotary evaporator. After that, the concentrated solutes were cleaned up by using 0.2 µm syringe filters and sodium sulfate (Na₂SO₄) cartridges to remove particles and water, respectively. The purified liquid extracts were concentrated to 1 mL under a gentle stream of nitrogen. 8:1 FA was added as an internal standard just prior to GC-MS analysis to correct sample volume variability. The other half of GFF was used to determine particle phase PFCAs. It was soaked in 10 mL methanol and shaken for 1 hour. The extraction process was performed for 4 cycles (~40 mL in total). Mass-labeled PFCAs (¹³C₂-PFHxA, ¹³C₄-PFOA and ¹³C₂-PFDA) were spiked into the samples

to calculate their recoveries. The extracts were passed through ENVI-carb filter (Supelco, U.S.A.) to eliminate matrix. Then, the purified extracts were evaporated, and reconstituted into HPLC-MS/MS mobile phase (40% acetonitrile in ultrapure water) to a final volume 1 mL.

Chemicals and Instrumental analysis: Table 2 shows analytical parameters of FTOHs and PFCAs. Ten target compounds; three FTOHs and seven PFCAs, and 8:1 FA (internal standard) were purchased from Wako Pure Chemicals (Osaka, Japan). Four mass-labeled ($^{13}\text{C}_2$ 8:2 FTOH, $^{13}\text{C}_2$ -PFHxA, $^{13}\text{C}_4$ -PFOA and $^{13}\text{C}_2$ -PFDA) were purchased from the Wellington Laboratory Inc. (Ontario, Canada). Analytical or HPLC grade were used for all solvents and reagents. FTOHs were analyzed by an Agilent 6890 gas chromatography-Agilent 5973 mass spectrometer in electron ionization mode using selective ionmonitoring, while PFCAs were measured by an Agilent 1200SL high performance liquid chromatography coupled with an Agilent 6400 Triple Quadrupole mass spectrometer in electrospray ionization negative mode using multiple reactions monitoring.

Table 1 Locations and characteristics of the sampling sites

Sampling sites		Locations	Site characteristics		
<i>Outdoor air-WWTs</i>			Capacity (m ³ /d)	No. of industries	Industrial types
1.	Industrial WWT1	Industrial estate1, at aeration tank	4,000	213	Chemical and related (55.4%), Petroleum and energy (19.2%), Plastic (7.0%)
2.	Industrial WWT2	Industrial estate2, at aeration tank	16,000	244	Automotive (32.9%), Steel and metal (17.5%), Electronics and electrical (14.0%)
3.	Industrial WWT3	Industrial estate3, near aeration tank	45,000	350	Steel and metal (15.0%), Paper and printing (13.0%), Chemical and related (12.0%)
4.	Domestic WWT4	Suburban area, at aeration tank	157,000	Service area 44 km ² , 418,000 people	
<i>Outdoor air-Residential areas</i>			No. of households	Population density (capita/km ²)	Characteristics
5.	Residential area1	Industrial area	42,295	390	Northeast, 5 km from industrial estate1
6.	Residential area2	Urban area	55,575	6,441	~1 km from the main road
7.	Residential area3	Urban area	51,686	16,098	~10 m from traffic lanes of the main road
8.	Residential area4	Suburban area	14,568	640	at the university
<i>Indoor air</i>		Room size (m ²)		Activities/ characteristics	
9.	Condominium	Suburban area	30	Recently built, painted and decorated (new furniture and carpets)	
10.	Printing shop	Urban area	48	Document and poster printing	
11.	Office1	Suburban area	12	Instructor's room in the university, re-painted room	
12.	Office2	Urban area	225	Documenting and report preparing	
13.	Laboratory	Suburban area	74	Water analysis laboratory in the university	
14.	Furniture shop	Suburban area	150	Wood furniture, wind through opened door, window and porous wall	

Table2 Analytical parameters of FTOHs (by GC-MS) and PFCAs (by HPLC-MS/MS)

Compounds	Abbreviation	Ion1 (m/z)	Ion2 (m/z)	RT (min)	MDL (pg/m ³)	MQL (pg/m ³)	Blank (n=4) (pg/m ³)
2-Perfluorohexyl ethanol	6:2 FTOH	95	31, 69, 131	9.0	10.3	34.2	<10.3
2-Perfluorooctyl ethanol	8:2 FTOH	95	31, 69, 131	10.3	9.5	31.7	<9.5
2-Perfluorodecyl ethanol	10:2 FTOH	95	31, 69, 131	11.7	11.2	37.5	<11.2
Perfluorohexanoic acid	PFHxA	313	269	2.8	0.06	0.11	<0.06
Perfluoroheptanoic acid	PFHpA	363	319	4.7	0.06	0.17	<0.06
Perfluorooctanoic acid	PFOA	413	369	7.2	0.06	0.17	<0.06
Perfluorononanoic acid	PFNA	463	419	9.9	0.06	0.11	<0.06
Perfluorodecanoic acid	PFDA	513	469	12.7	0.06	0.22	<0.06
Perfluoroundecanoic acid	PFUnDA	563	519	15.4	0.39	1.22	<0.39
Perfluorododecanoic acid	PFDoDA	613	569	18.0	0.39	1.22	<0.39
1H,1H-Perfluoro-1-nonanol	8:1 FA	69	31, 95, 131	10.5	7.0	23.2	-
2-Perfluorooctyl-[1,1- ² H ₂]-[1,2- ¹³ C ₂]-ethanol	¹³ C ₂ -8:2 FTOH	131	31, 69, 95	10.3	17.1	57.0	-
Perfluoro-n-[1,2- ¹³ C ₂]hexanoic acid	¹³ C ₂ -PFHxA	315	271	2.8	0.06	0.11	-
Perfluoro-n-[1,2,3,4- ¹³ C ₄]octanoic acid	¹³ C ₄ -PFOA	417	373	7.2	0.06	0.17	-
Perfluoro-n-[1,2- ¹³ C ₂]decanoic acid	¹³ C ₂ -PFDA	515	471	12.7	0.06	0.17	-

Note: Ion1= quantification ion (GC-MS) and parent ion (HPLC-MS/MS), Ion2 = confirmation ion (GC-MS) and daughter ion (HPLC-MS/MS), RT = Retention time, MDL = Method detection limit, MQL = Method quantification limit

Calibration and Validation: The concentrations were calculated using five points standard curve analysis for FTOHs (10-1,000 pg/ μ L), and PFCAs (0.1-10 pg/ μ L), in which the determination coefficients (R^2) were more than 0.99. Instrument detection limits (*IDLs*) were defined as concentrations with signal to noise ratio (*S/N*) equal to 3:1. Instrument quantification limits (*IQLs*) were used for quantifying analytes, which were defined by *S/N* 10:1. *MDLs* and *MQLs* were calculated from the *IDLs* and *IQLs*, respectively, which expressed as concentrations by dividing by an air sample volume of 720 m³. FTOHs and PFCAs in field blanks were lower than *MDLs*, indicating no contamination during the process. PFDoDA was not detected in any samples, so that it was excluded in the next discussion. The recovery of ¹³C₂-8:2 FTOH was 61-119% and 73-132% for particle and gas phases, respectively. For PFCAs, the range of recovery rate of ¹³C₂-PFHxA, ¹³C₄-PFOA and ¹³C₂-PFDA were 71-109%, 73-111% and 92-125%, respectively.

Results and discussion

FTOHs and PFCAs were detected from all indoor and outdoor air sampling sites in Thailand (**Fig.1**), which indicated their contamination in the study area. Total concentration of three FTOHs ranged from 1,690 to 13,030 pg/m³, while total concentration of six PFCAs were 4-110 pg/m³ which were two orders of magnitude lower than FTOHs (**Fig.2**). The highest outdoor FTOHs and PFCAs concentrations were found at an aeration tank of the industrial WWTP1. Wastewater might contain FTOHs and PFCAs, and they could be released to the air by aeration process, which was reported by a previous study⁹. High FTOHs concentrations (7,260-13,030 pg/m³) were also detected in condominium, printing shop, office1 and office2, which were higher than outdoor residential areas (1,830-3,020 pg/m³). FTOHs are used as surfactants in several products such as carpet, textile, paper, paints, coatings and adhesives, and there is potential for residual FTOHs (unreacted and unbound) to be released from these products¹⁷. High concentrations of FTOHs in such kind of indoor places were also reported by a previous study¹⁸. FTOHs were mainly distributed in gas phase fractions (89-100%). However, composition profiles of FTOHs and PFCAs were different in each sampling site. 10:2 FTOH (26-76%) was the dominant compound in most samples, followed by 8:2 FTOH (14-55%) and 6:2 FTOH (5-48%). For PFCAs, PFOA (9-72%) and PFHxA (3-50%) were dominant among the other compounds of PFCAs. This different distribution patterns might be caused by the variety of their emission sources in the measurement areas.

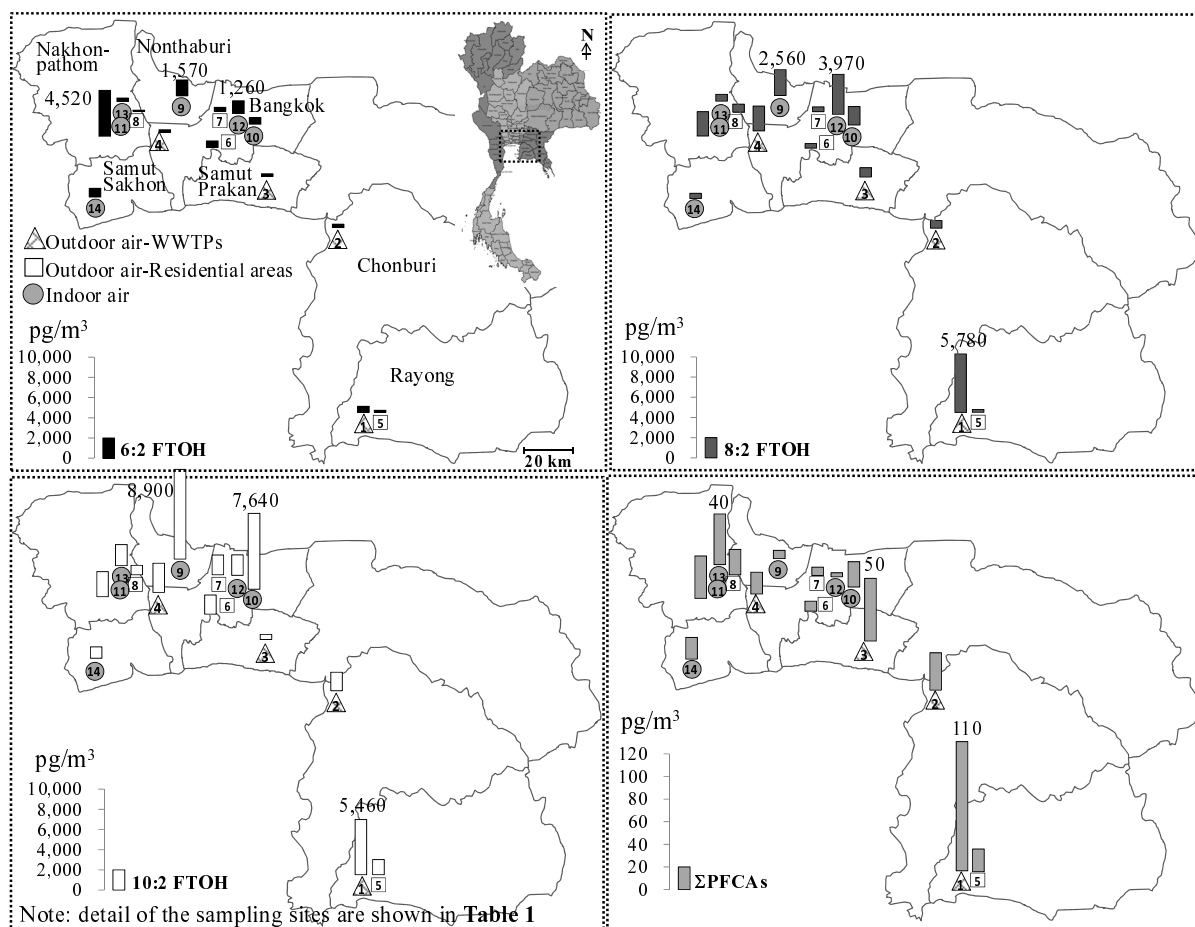


Figure 1 Distribution of FTOHs (gas and particle phases) and Σ PFCA s (particle phase) in the study areas

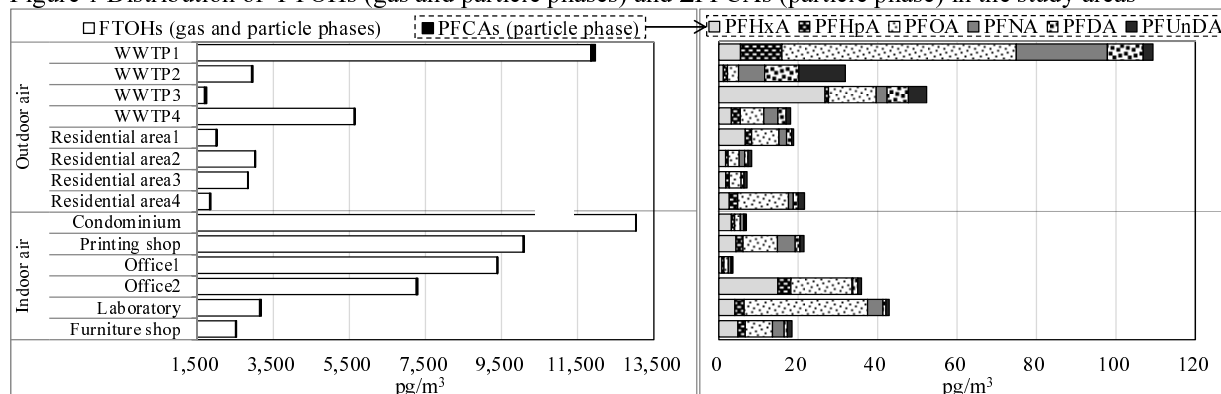


Figure 2 Concentration of FTOHs and PFCA s in outdoor and indoor air in Thailand

Compared to the studies in other countries (Fig. 3), outdoor FTOHs concentrations in Thailand were found at higher levels in most sampling sites, while the indoor concentrations were found at similar high levels as the others. Outdoor and indoor PFCA s concentrations in this study were about two orders of magnitude lower than FTOHs, which were also reported in most of the mentioned studies. Therefore, high concentration levels of airborne FTOHs might have high possibility to contribute to PFCA s contamination worldwide. To confirm this hypothesis, it is needed to investigate deposition and degradation pathways of FTOHs in the air to PFCA s in water environment.

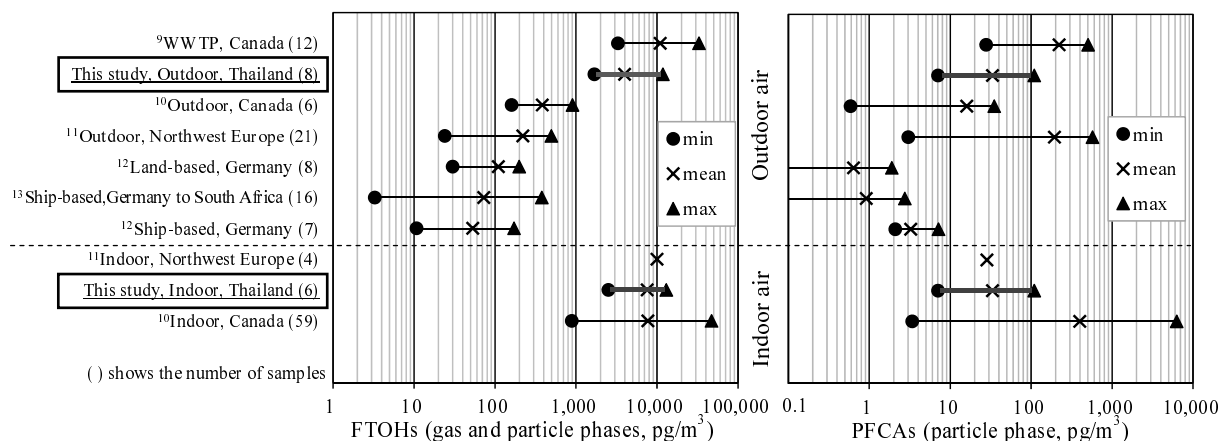


Figure 3 Comparison of FTOHs and PFCAs in outdoor and indoor air in Thailand with other countries

Acknowledgements

The study was supported by JSPS (22360214, 25289169) and Environmental Management Leadership program, Kyoto University.

References

- Ahrens L. (2011); *J. Environ. Monit.* **13**: 20-31
- Andersen ME, Butenhoff JL, Chang SC, Farrar DG, Kenedy GL, Lau C, Olsen GW, Seed J, Wallace KB. (2008); *J. Toxicol. Sci.* **102**(1): 3-14
- Conder JM, Hoke RA, Wolf WD, Russel MH, Buck RC. (2007); *Environ. Sci. Technol.* **42**(4): 995-1003
- Fujii S, Polprasert C, Tanaka S, Lien NPH, Qiu Y. (2007); *Water Supply and Technology-AQUA*. **56.5**:313-26
- Lau C, Anitole K, Hodes C, Lai D, Hutchens AP, Seed J. (2007); *J. Toxicol. Sci.* **99**(2): 366-94
- Barton CA, Kaiser MA, and Russell MH. (2007); *J. Environ. Monit.* **9**: 839-46
- Ellis DA, Martin JW, De Silva AO, Mabury SA, Hurley MD, Andersen MPS, Wallington TJ. (2004); *Environ. Sci. Technol.* **38**: 3316-21
- Butt CM, Muir DCG, Mabury SA. (2010); *Environ. Sci. Technol.* **44**(13): 4973-80
- Vierke L, Ahrens L, Shoeib M, Reiner EJ, Guo R, Palm WU, Ebinghaus R, Harner T. (2011); *Environ. Chem.* **8**: 363-71
- Shoeib M, Harner T, Webster GM, Lee SC. (2011); *Environ. Sci. Technol.* **45**: 7999-8005
- Barber JL, Berger U, Chaemfa C, Huber S, Jahnke A, Temme C, Jones KC. (2007); *J. Environ. Monit.* **9**: 530-41
- Dreyer A, Ebinghaus R. (2009); *Atmos. Environ.* **43**: 1527-35
- Jahnke A, Berger U, Ebinghaus R, Temme C. (2007); *Environ. Sci. Technol.* **41**(9): 3055-61
- Dreyer A, Weinberg I, Temme C, Eribghaus R. (2009); *Environ. Sci. Technol.* **43**(17): 6507-14
- Kunacheva C, Fujii S, Tanaka S, Boontanon SK, Musirat C, Wongwatthana T, Shivakoti BR. (2009); *Organohalog. Compd.* **71**: 216-21
- Chularueangakorn P, Tanaka S, Kunacheva C, Fujii S, Harada H, Boontanon SK. (2012); *Environ. Eng. Res.* **68**(7): III341-9
- Dinglasan-PMJA and Mabury SA. (2006); *Environ. Sci. Technol.* **40**(5): 1447-53
- Langer V, Dreyer A, and Ebinghaus R. (2010); *Environ. Sci. Technol.* **44**(21): 8075-81