### *IN VIVO* METABOLISM OF 2,2',4',5,5'-PENTACHLOROBIPHENYL AND ITS INDUCTIVE ACTIVITY OF DRUG-METABOLIZING ENZYMES : SPECIES DIFFERENCE BETWEEN RATS AND MICE

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#### Introduction

A number of the methylsulfonyl (MeSO<sub>2</sub>) metabolites of polychlorinated biphenyls (PCBs) have been found in several species of animals in Canada and Sweden<sup>14</sup> and in healthy humans<sup>5-8</sup>.

We previously reported that nine 3-MeSO<sub>2</sub> metabolites of PCBs were the potent inducers of hepatic microsomal drug-metabolizing enzymes in rats<sup>9-11</sup>. Additionally we showed that the seven 3-MeSO<sub>2</sub> and two 4-MeSO<sub>2</sub> metabolites of tetra-, penta- and hexachlorobiphenyls (tetra-, penta- and hexaCBs) reduced the level of serum thyroxine<sup>12-14</sup>, suggesting that the metabolites may act as endocrine-disrupters. Recently we indicated that increase in the hepatic T<sub>4</sub> glucuronidation through the seven 3-MeSO<sub>2</sub> and one 4-MeSO<sub>2</sub> metabolites-mediated induction of both UGT1A1 and UGT1A6 caused the reduction of serum T<sub>4</sub> levels<sup>15</sup>. These previous findings<sup>9-15</sup> indicated that the metabolites, mainly MeSO<sub>2</sub> metabolites, contributed to the biological effects such as endocrine-disruption and induction of drug-metabolizing enzymes by PCB congeners in several mammalian species.

Several mammalian species show different responses to PCB-derived toxicity such as endocrinedisruption, drug-metabolizing enzyme induction and so on<sup>16</sup>. The difference might be attributed to the species differences in the metabolism pattern of PCB congeners and/or the induction of drugmetabolizing enzymes by the chemicals.

In this study, we examined the species differences in the *in vivo* metabolism of 2,2',4',5,5'-pentaCB, which is one of major MeSO<sub>2</sub> metabolites accumulated in blubber of seals in the Baltic', and in the induction of drug-metabolizing enzymes by the compound between rats and mice.

### **Materials and Methods**

**Chemicals.** 2,2',4',5,5'-pentaCB was synthesized by using the Cadogan coupling reactions<sup>17</sup>. 3- and 4-MeS, 3- and 4-MeSO, and 3- and 4-MeSO<sub>2</sub>-2,2',4',5,5'-pentaCBs were prepared by the method as described before<sup>18</sup>. The purity of these compounds was >99% when analyzed by gas chromatography (GC). All other chemicals used in the present experiments were commercially obtained.

Animal treatments. Male Wistar rats, weighing 180-200 g, and male ddy mice, weighing 27-35 g, were housed three or four per cage in the laboratory with free access to commercial chow and tap water, and maintained on a 12-hr dark/light cycle (8:00 A.M.-8:00 P.M. light) in a room with controlled temperature ( $24.5 \pm 1 \Box$ ) and humidity ( $55 \pm 5\%$ ). Rats and mice received an intraperitoneal injection of 2,2',4',5,5'-pentaCB ( $342 \mu mol/kg$ ) dissolved in Panacete 810 (5 ml/kg). Control animals received an equivalent volume of vehicle. All animals were starved for 18 hr prior to killing by decapitation at designated time after dosing.

**Preparation of hepatic microsomes and soluble fractions, and enzyme assays.** Microsomes and soluble fractions (105,000g supernatant) were prepared according to the procedure described previously<sup>9</sup>. The protein content was determined by the method of Lowry *et al.*<sup>19</sup> with bovine serum albumin as a standard. Total cytochrome P450 content was estimated according to the method of Omura and Sato<sup>20</sup>. The activity of alkoxyresorufin O-dealkylase in microsomes was determined by the method of Burke *et al.*<sup>21</sup>. The microsomal activities of UDP-glucuronosyltransferase (UDP-GT) toward 4-nitrophenol and chloramphenicol were determined as described by Isselbacher *et al.*<sup>22</sup> and Ishii *et al.*<sup>23</sup>, respectively. The glutathione S-transferase (GST) activities were measured using four different substrates, 1-chloro-2,4-dinitrobenzene (CDNB), 1,2-dichloro-4-nitrobenzene (DCNB), ethacrynic acid (EA) and 1,2-epoxy-3-(p-nitrophenoxy)propane (EPNP) by the method of Habig *et al.*<sup>24</sup>

**Determination of 2,2',4',5,5'-pentaCB and its sulfur containing metabolites in the liver and feces.** The concentrations of 2,2',4',5,5'-pentaCB and its sulfur containing metabolites present in the liver and feces were determined with GC as described priviously<sup>2</sup>.

#### **Results and Discussion**

The elimination rate of 2,2',4',5,5'-pentaCB and the concentrations of 3- and 4-MeSO<sub>2</sub> metabolites in the liver for 8 days after the administration of 2,2',4',5,5'-pentaCB were different between rats and mice (Fig. 1). The fecal excretion of 3-MeS metabolite during 8 days accounted for 0.3 and 0.6% of the dose of 2,2',4',5,5'-pentaCB in rats and mice, respectively. On the other hand, the fecal excretion of 4-MeS metabolite accounted for 2 and 1.3% of the dosage in rats and mice, respectively. The level of cumulative excretion of 3- and 4-MeSO<sub>2</sub> metabolites during 8 days was 5-15 times higher in mice than in rats.

2,2',4',5,5'-pentaCB administration resulted in the increase in hepatic benzyloxyresorufin O-dealkylase activity: 34- and 11-fold, pentoxyresorufin O-dealkylase activity (CYP2B1/2): 16- and 6-fold, and ethoxyresorufin O-dealkylase activity (CYP1A1/2): 1.7- and 1.3-fold in rats and mice, respectively, at 8 days after the administration (Fig. 2). The administration of 2,2',4',5,5'-pentaCB extensively increased the UDP-GT (UGT2B1) activity toward chloramphenicol in rats, and slightly increased that in mice. No increase in UDP-GT (UGT1A6) activity toward 4-nitrophenol by 2,2',4',5,5'-pentaCB was observed in both species. In addition, the species difference was observed in the alteration of GST activities using four substrates after the administration of 2,2',4',5,5'-pentaCB.

In conclusion, there are marked differences in the elimination and metabolism of 2,2',4',5,5'-pentaCB and in induction pattern of phase I microsomal drug-metabolizing enzymes, UDP-GT and GST by 2,2',4',5,5'-pentaCB between rats and mice. These differences might be correlated with those in the susceptibility of animals to PCBs-derived toxicity.

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Fig. 1. Time courses of liver concentrations of 2,2',4',5,5'-pentaCB, and its 3- and 4-MeSO<sub>2</sub> metabolites after the administration of 2,2',4',5,5'-pentaCB to rats and mice. Animals were given 2,2',4',5,5'-pentaCB (342  $\mu$ mol/kg) i.p. and killed at the indicated times after the administration. Each point represents single values and the mean ± S.E. (vertical bars) for two to three animals. \**P*<0.05, significantly different from the corresponding value of rats. (0) Rats, (•) mice.



Fig. 2. Effects of 2,2',4',5,5'-pentaCB on the activities of drug-metabolizing enzymes of hepatic microsomes in rats and mice. The experimental conditions were the same as described in the note to Fig. 1. Each point represents the mean  $\pm$  S.E. (vertical bars) for three to four animals. \**P*<0.05, significantly different from control (0 day). (0) Rats, (•) mice.

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### References

- Haraguchi K., Athanasiadou M., Bergman Å., Hovander L. and Jensen S. (1992) Ambio 21, 546-549.
- Bergman Å., Athanasiadou M., Bergek S., Haraguchi K., Jensen S. and Klasson-Wehler E. (1992) Ambio 21, 570-576.
- 3. Bergman Å., Norstrom R.J., Haraguchi K., Kuroki H. and Béland P. (1994) Environ. Toxicol. Chem. 13, 121-128.
- 4. Letcher R.J., Norstrom R.J. and Bergman Å. (1995) Sci. Total Environ. 160/161, 409-420.
- 5. Haraguchi K., Kuroki H. and Masuda Y. (1986) J. Chromatogr. 361, 239-252.
- 6. Haraguchi K., Kuroki H. and Masuda Y. (1989) Chemosphere 18, 477-484.
- 7. Norén K., Lundén Å., Pettersson E. and Bergman Å. (1996) Environ. Health Perspect. 104, 766-772.
- 8. Weistrand C. and Norén K. (1997) Environ. Health Perspect. 105, 644-649.
- 9. Kato Y., Haraguchi K., Kawashima M., Yamada S., Masuda Y. and Kimura R. (1995) Chem.-Biol. Interact. 95, 257-268.
- 10. Kato Y., Haraguchi K., Kawashima M., Yamada S., Isogai M., Masuda Y. and Kimura R. (1995) *Chem.-Biol. Interact.* 95, 269-278.
- 11. Kato Y., Haraguchi K., Tomiyasu K., Saito H., Isogai M., Masuda Y. and Kimura R. (1997) Environ. Toxicol. Pharmacol. 3, 137-144.
- Kato Y., Haraguchi K., Shibahara T., Masuda Y. and Kimura R. (1998) Arch. Toxicol. 72, 541-544.
- 13. Kato Y., Haraguchi K., Shibahara T., Yumoto S., Masuda Y. and Kimura R. (1999) *Toxicol. Sci.* 48, 51-54.
- Kato Y., Haraguchi K., Shibahara T., Yumoto S., Masuda Y. and Kimura R. (2000) Chemosphere 40, 1233-1240.
- 15. Kato Y., Haraguchi K., Shibahara T., Shinmura Y., Masuda Y. and Kimura R. (2000) Chem.-Biol. Interact. 125, 107-115.
- 16. Safe S.H. (1994) Crit. Rev. Toxicol. 24, 87-149.
- 17. Cadogan J.I.G. (1962) J. Chem. Soc. 4257-4258.
- 18. Haraguchi K., Kuroki H. and Masuda Y. (1987) J. Agric. Food Chem. 35, 178-182.
- 19. Lowry O.H., Rosebrough N.J., Farr A.L. and Randall R.J. (1951) J. Biol. Chem. 193, 265-275.
- 20. Omura T. and Sato R. (1964) J. Biol. Chem. 239, 2370-2378.
- 21. Burke M.D., Thompson S., Elcombe C.R., Halpert J., Haaparanta T. and Mayer R.T. (1985) Biochem. Pharmacol. 34, 3337-3345.
- 22. Isselbacher K.J., Chrabas M.F. and Quinn R.C. (1962) J. Biol. Chem. 237, 3033-3036.
- 23. Ishii Y., Tsuruda K., Tanaka M. and Oguri K. (1994) Arch. Biochem. Biophys. 315, 345-351.
- 24. Habig W.H., Pabst M.J. and Jakoby W.B. (1974) J. Biol. Chem. 249, 7130-7139.